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Synthesis of Fluorescence-Labelled Glycosidic Prodrugs Based on the Cytotoxic Antibiotic Duocarmycin

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Dedicated to Professor Carmen Najera on the occasion of her 60th birthday

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The synthesis of the glycosidic prodrugs (1S)-30a, (1S,10R)-**30b** and (1S,10R)-**32** labelled with different fluorescence dyes at different positions at the aromatic A-ring in 2 is described; the compounds are structurally based on the cytotoxic antibiotic duocarmycin SA. For binding, the amino compounds (1S)-3a and (1S,10R)-3b were treated with the commercially available succinimides of the dyes 5-SFX (29) and D10162 (31), respectively.

Introduction

Over the last few years we have successfully developed novel glycosidic prodrugs^[1] such as **2a** and **2b** (Scheme 1), which can be used for selective treatment of cancer through antibody-directed enzyme prodrug therapy (ADEPT).^[2,3]

The compounds are based on the cytotoxic antibiotic duocarmycin SA (1, IC₅₀ \approx 10 pm, L1210)^[4] and have as yet unsurpassed selectivity factors of $QIC_{50} = 3200$ and 4800, respectively (QIC₅₀ = IC₅₀ of prodrug/IC₅₀ of prodrug in the presence of the cleaving enzyme).^[5] Here we describe the synthesis of the fluorescence-labelled derivatives (1S)-**30a**, (1*S*,10*R*)-**30b** and (1*S*,10*R*)-**32**, in order to determine their cellular uptakes and mode(s) of action. Fluorescencelabelled compounds can thus be used for verification of their interaction with cellular targets at the molecular level either by confocal laser scanning microscopy for cell cultures^[6] or with the aid of the Explore-Optics instrument (GE) for mice.^[7]

For binding to the fluorescence dyes we introduced aminomethyl groups into 2a and 2b at different positions in the A-ring. This should also allow us to determine the influence of the binding position on the activities of the labelled com-

$$\mathsf{MeO_2C} \xrightarrow{\mathsf{N}} \overset{\mathsf{N}}{\mathsf{N}} \overset{\mathsf{N}}{\mathsf{N}} \overset{\mathsf{OMe}}{\mathsf{OMe}}$$

1 (+)-Duocarmycin SA

$$\begin{array}{lll} \text{(1S)-2a} & & R^1 = \ R^2 = \ R^3 = H \\ \text{(1S,10}R\text{)-2b} & & R^1 = \ R^2 = H, \ R^3 = CH_3 \\ \text{(1S)-3a} & & R^1 = CH_2NH_2, \ R^2 = \ R^3 = H \\ \text{(1S,10}R\text{)-3b} & & R^1 = H, \ R^2 = CH_2NH_2, \ R^3 = CH_3 \\ \end{array}$$

Scheme 1. Structures of (+)-duocarmycin SA (1) and the glycosidic prodrugs 2a and 2b, as well as the compounds 3a and 3b, each containing an aminomethyl group for binding with the fluorescence

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pounds. The synthesis of the corresponding substrates 3a and 3b started from meta-diiodobenzene and para-diiodobenzene, respectively. As dyes, flurorescein (4, Scheme 2) and dapoxyl (5) were used, through application of the commercially available activated compounds 5-SFX (29, Scheme 11, below) and D10162 (31, Scheme 13, below).

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These show absorption maxima of $\lambda_{\rm max} = 494$ nm for **4** and $\lambda_{\rm max} = 395$ nm for **5** and emission maxima at $\lambda_{\rm max} = 521$ nm for **4** and $\lambda_{\rm max} = 574$ nm for **5** with high extinction coefficients.

Scheme 2. Structures of fluorescein (4) and dapoxyl (5).

Results and Discussion

Formylation of the diiodobenzenes **6a** and **6b** (Scheme 3) by halogen/metal exchange with *n*-butyllithium at –78 °C in THF, followed by addition of DMF, led to the iodobenzal-dehydes **7a** and **7b**, respectively. The aldehydes were then converted into the styrene derivatives **8a** and **8b** through Wittig–Horner reactions with the phosphonate **9** and NaH in THF. Acid-catalysed deprotection of **8a** and **8b** in TFA/H₂O yielded the carboxylic acids **10a** and **10b**, which were transformed into the naphthalene derivatives **11a** and **11b** through intramolecular Friedel–Crafts reactions by treatment with acetic anhydride and KOAc under reflux. In the

$$R^{1} = I, R^{2} = H$$

$$6a R^{1} = I, R^{2} = H$$

$$6b R^{1} = H, R^{2} = I$$

$$R^{2} = H$$

$$R$$

Scheme 3. Synthesis of the naphthalenes **11a** and **11b** starting from the diiodobenzenes **6a** and **6b**, respectively. Reagents and conditions: (a) nBuLi, THF, -78 °C, 20 min, then DMF, r.t., 2 h; 84–95%; (b) **9**, NaH, THF, 0 °C \rightarrow r.t., 19 h; 63-72%; (c) TFA/H₂O (9:1), r.t., 6 h, 70-100%; (d) KOAc, Ac₂O, reflux, 1 h, 87-90%.

case of **10a** we obtained a mixture of the desired **11a** and its regioisomer **11c** in a ratio of 2.5:1 (determined by ¹H NMR spectroscopy); at this stage it was not possible to separate the two compounds. In contrast, when starting from **10b**, compound **11b** was formed as the only product.

The acetates 11a/c and 11b were solvolysed with K_2CO_3 in ethanol to yield the phenols 12a/c and 12b (Scheme 4), which were protected at their phenolic hydroxy groups by treatment with BnBr and catalytic amounts of TBAI to give 13a/c and 13b. Exchange of the iodide for CN with CuCN under microwave conditions led to 14a/c and 14b.^[8]

R¹
$$CO_2Et$$

R² R^3 OAc

11a R¹ = I, R² = H, R³ = H

11b R¹ = H, R² = H, R³ = I

11c R¹ = H, R² = I, R³ = H

12b R¹ = H, R² = H, R³ = I

11c R¹ = H, R² = I, R³ = H

CO₂Et

C

R³ OH

12a R¹ = I, R² = H, R³ = H

12b R¹ = H, R² = H, R³ = H

12c R¹ = H, R² = I, R³ = H

b

CO₂Et

R³ OH

12a R¹ = I, R² = H, R³ = H

12b R¹ = H, R² = I, R³ = H

13c R¹ = H, R² = H, R³ = H

13c R¹ = H, R² = H, R³ = H

13c R¹ = H, R² = I, R³ = H

13c R¹ = H, R² = I, R³ = H

13c R¹ = H, R² = I, R³ = H

13c R¹ = H, R² = I, R³ = H

13c R¹ = H, R² = I, R³ = H

13c R¹ = H, R² = I, R³ = H

14d R¹ = CN, R² = H

15b R¹ = H, R² = CN

Scheme 4. Synthesis of **16a** and **16b** starting from the naphthalene derivatives **11a** and **11b**, respectively. Reagents and conditions: (a) K₂CO₃, EtOH, reflux, 4 h, 84–92%; (b) BnBr, K₂CO₃, TBAI, DMF, r.t., 15 h, 82–94%; (c) CuCN, DMF, 140 °C (microwaves), 1 h, 92–94%; (d) LiOH·H₂O, THF/MeOH/H₂O, r.t., 15 h, 87–98%; (e) DPPA, NEt₃, *t*BuOH, molecular sieves (4 Å), reflux, 20 h, 95–100%.

At this stage it was possible to recrystallise the **14a/c** mixture from *n*-pentane/EtOAc to afford pure **14a**. Hydrolysis of the ethyl carboxylates **14a** and **14b** with lithium hydroxide, followed by formation of the corresponding acyl azides and Curtius rearrangements with use of diphenylphosphoryl azide in *tert*-butyl alcohol, led to the Boc-protected aminonaphthalenes **16a** and **16b** (Scheme 4).

Bromination of **16a** and **16b** with NBS and catalytic amounts of sulfuric acid gave the compounds **17a** and **17b** (Scheme 5), whereas iodination of **16a** mediated by NIS and catalytic amounts of *p*-toluenesulfonic acid yielded compound **17c**.

For the synthesis of the aminomethyl-substituted benzoindoles **3a** and **3b** from **17** we envisaged two different approaches (Scheme 6): firstly, *N*-alkylation with the chloroallyl chlorides **18a** and **18b**, respectively, followed by radical



R1 NHBoc

$$R^2$$
 OBn OBn $R^2 = H$ $R^1 = CN, R^2 = H, X = Br$ $R^1 = CN, R^2 = H, X = Br$ $R^1 = CN, R^2 = H, X = Br$ $R^1 = CN, R^2 = H, X = Br$ $R^2 = CN, X = Br$ $R^2 = CN, X = Br$ $R^2 = CN, R^2 = H, X = Br$ $R^2 = CN, R^2 = H, X = Br$ $R^2 = CN, R^2 = H, X = Br$ $R^2 = CN, R^2 = H, X = Br$ $R^2 = CN, R^2 = H, X = Br$ $R^2 = CN, R^2 = H, X = Br$

Scheme 5. Synthesis of 17a–17c. Reagents and conditions: for 17a/17b: NBS, cat. H_2SO_4 , THF, -78 °C, 4 h, 81–94%; for 17c: NIS, cat. TsOH· H_2O , THF, 50 °C, 1.5 h, 81%.

cyclisation, and secondly, N-alkylation with the epoxy nosylates (2R)-**20a** and (2R,3R)-**20b**, respectively, followed by metal-induced cyclisation. The latter approach has the advantage of leading to enantiopure – and in the case of (2R,3R)-**20b** also diastereopure – product(s).

N-Alkylation of 17c and 17b with the allyl halides 18a and 18b led to 19a and 19b, whereas treatment of 17a and 17b with the epoxy nosylates (2R)-20a and (2R,3R)-20b gave (2R)-21a and (2R,3R)-21b. Interestingly, compounds of type 21 each contain an axis of chirality along the naphthyl–N bond, so they are obtained as mixtures of diastereomers. [9]

Radical cyclisation of **19a** and **19b**, mediated by tris(trimethylsilyl)silane and azobisisobutyronitrile in toluene, worked well to afford the cyclised products **22a–c** (Scheme 7) in 85–92% yields as racemic mixtures, and in the case of **19b** in a nearly 1:1 ratio of the two possible diastereomers **22b/22c**.^[10] Similarly, metal-induced cyclisation of (2*R*,3*R*)-**21b** in the presence of ZnCl₂, MeLi and TMS-SCN led to the benzindole derivative (1*S*,10*R*)-**23b** in 53% yield, in this case as a pure enantiomer (>99% *ee*) and a pure diastereomer. In comparison, (2*R*)-**21a** gave the

Scheme 6. Synthesis of **19** and **21** from **17**. Reagents and conditions: (a) NaH, DMF, **18**, r.t., 4–14 h, 77–93%; (b) NaH, DMF, **20**, r.t., 21 h, 88–89%.

Scheme 7. Synthesis of *rac-***22** and enantiopure **23**. Reagents and conditions: (a) TTMSS, AIBN, toluene, 80 °C, 4.5 h, 85–92%; (b) (2*R*)-**20a**: *n*BuLi (3.0 equiv.), -85 °C, 1 h, 12%; (2*R*,3*R*)-**20b**: ZnCl₂ (1.5 equiv.), MeLi (6.0 equiv.), TMS-SCN (1.5 equiv.), 55%.

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Scheme 8. Synthesis of the phenols **24**. Reagents and conditions: (a) (1*S*)-**24a**: Pd/C, H₂, EtOAc, r.t., 5 h, 90%; (1*S*,10*R*)-**24b**: Pd/C, NH₄HCO₂, THF, 40 °C, 25 min, 86% (transfer hydrogenation).

desired product (1S)-23a in only 12% yield on treatment with nBuLi at -85 °C. All other attempts to initiate the cyclisation via a zincate or cuprate failed.

Treatment of (1S)-23a under Appel conditions afforded enantiopure (1S)-22a, [10] whereas treatment of (1S,10R)-23b with SOCl₂ in the presence of pyridine led to enantiopure (1S,10R)-22b. Alternatively, the enantiomeric mixture of rac-22a obtained from 19a could be resolved by preparative HPLC on a chiral stationary support (Chiralpak IA), with ee values of 99.7% for (+)-(1R)-22a and 99.2% for (-)-(1S)-22a. The two diastereomers rac-22b and rac-22c obtained from 19b were separated by column chromatography on silica gel, whereas rac-22b was resolved again by preparative HPLC on chiral support (Chiralpak IA) to obtain the desired (+)-(1S,10R)-22b with an ee value of 99.9%. Neither the diastereomer 22c nor the enantiomer (-)-(1R,10S)-22b were included in further investigations, because we have shown that the corresponding seco drugs – without the cyano group – have much lower cytotoxicities.

Deprotection of **22a** and **22b** either with Pd/C under H_2 or with ammonium formate, respectively, gave the free phenols **24a** and **24b** in good yields (Scheme 8).

Next, the galactose moiety and the DNA binder were introduced (Scheme 9). For this purpose (1S)-24a and (1S,10R)-24b were glycosylated with the galactose trichloroacetimidate 25 in the presence of BF₃·OEt₂ at -10 °C. Addition of an excess of BF₃·OEt₂ at room temp. led to the corresponding secondary amine as a result of Boc removal. This amine was then coupled with the DNA binder DMAI 26 without isolation in the presence of EDC·HCl as the activating agent.

Nitrile reduction in (1*S*)-27a and (1*S*,10*R*)-27b in accordance with the method of Wrobel et al.^[11] with PtO₂·H₂O under H₂ afforded the corresponding primary amines 28a and 28b (Scheme 10), and these were deprotected under basic conditions with NaOMe in MeOH to give the desired aminomethyl-substituted prodrugs (1*S*)-3a and (1*S*,10*R*)-3b.

The final step in the synthesis of the fluorescence-labelled prodrugs was the coupling of (1S)-3a and (1S,10R)-3b with the activated commercially available dyes 5-SFX (29) as a derivative of fluorescein (4) and D10162 (31) as a derivative of dapoxyl (5). Coupling of (1S)-3a with 29 in DMF and Hünig base (Scheme 11), followed by purification by pre-

Scheme 9. Synthesis of the galactosides **27a** and **27b**. Conditions and reagents: (a) **26**, 0.5 equiv. BF₃·OEt₂, CH₂Cl₂, molecular sieves (4 Å), -10 °C, 3.5 h, then 3 equiv. BF₃·OEt₂, r.t., 6 h, **27**, EDC·HCl, DMF, r.t., 18 h, 52–53%.

parative HPLC (Kromasil 100 C18), gave the fluorescence-labelled prodrug (1*S*)-30a in 51% yield.

In the same manner, coupling of (1S,10R)-3b with 29 (Scheme 12) afforded the fluorescence-labelled prodrug (1S,10R)-30b in 88% yield.

In addition, the aminomethyl-substituted prodrug (1S,10R)-3b was also coupled with the dapoxyl derivative 31 under the same conditions as described for 29 to give the fluorescence-labelled prodrug (1S,10R)-32 in 89% yield (Scheme 13).



Scheme 10. Synthesis of the aminomethyl-substituted prodrugs 3. Reactions and conditions: (a) HCl/EtOH, PtO₂·H₂O, H₂, r.t., 48 h, 66–81%; (b) NaOMe/MeOH, MeOH, r.t., 30 min, prep. HPLC (Kromasil 100 C18), 70–95%.

$$H_2$$
N H_2 N H_2 N H_3 N H_4 H_4

Scheme 11. Coupling of the prodrug (1S)-3a with the NHS ester 29 to give (1S)-30a. Reagents and conditions: (a) iPr₂NEt, DMF, r.t., 18 h, prep. HPLC (Kromasil 100 C18), 51%.

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Scheme 12. Coupling of the prodrug (1S,10R)-3b with the NHS ester 29 to give (1S,10R)-30b. Reagents and conditions: (a) iPr_2NEt , DMF, r.t., 18 h, prep. HPLC (Kromasil 100 C18), 88%.

Scheme 13. Coupling of (1S,10R)-3b with 31 to give (1S,10R)-32. Conditions and reagents: (a) iPr_2NEt , DMF, r.t., 9 h, prep. HPLC, 89%.



Conclusions

We have prepared the fluorescence-labelled prodrugs (1S)-30a, (1S,10R)-30b and (1S,10R)-32 to allow investigations of their cellular uptakes and their mode(s) of action by confocal laser scanning microscopy in the case of cell cultures and with the Explore Optix (GE) instrument in the case of mice. The results will be published in due course in a more biologically orientated journal.

Experimental Section

General: All reactions were performed in flame-dried glassware under argon. Commercially available reagents were used without further purification. Thin layer chromatography (TLC) was carried out on precoated Alugram SIL G/UV254 (0.25 mm) plates from Macherey-Nagel & Co. Column chromatography (CC) was carried out on silica gel 60 from Merck with particle size 0.063–0.200 mm for normal-pressure and flash chromatography. IR spectra were determined with a Bruker Vektor 22 instrument, UV/Vis spectra with a Perkin-Elmer Lambda 2, and mass spectra with a Finnigan MAT 95 for EI HRMS and a Bruker Apex IV Fourier transform ion cyclotron resonance mass spectrometer for ESI HRMS. 1H NMR spectra were recorded variously with a Varian UNITY 300 MHz, a Varian Inova 500 MHz or a Varian Inova 600 MHz instrument. ¹³C NMR spectra were recorded at 75, 125, or 150 MHz. Spectra were taken at room temperature except when stated otherwise in deuterated solvents as indicated with use of the solvent peak as internal standard. The purities and stabilities of the target compounds were checked by HPLC-MS [ESI mass spectrometry with an ion-trap mass spectrometer LCQ (Finnigan)]. The columns used were Phenomenex Synergi Max-RP C12 $(150 \times 2 \text{ mm}, \text{ particle size } 4 \,\mu\text{m})$. Chromatographic separation by HPLC was performed with use either of a Kromasil 100 C18 column (250 × 20 mm, particle size 7 μm) or of a Chiralpak IA column (250 \times 20 mm, particle size 5 μ m).

(E/Z)-2-Amino-4-(benzyloxy)-N-(tert-butoxycarbonyl)-N-(3-chloroprop-2-enyl)-7-cyano-1-iodonaphthalene (19a): NaH (399 mg, 60% dispersion in mineral oil, 10.0 mmol, 2.5 equiv.) was added at room temperature to a solution of 17c (2.00 g, 4.00 mmol, 1.0 equiv.) in anhydrous DMF (50 mL), and the mixture was stirred for 1.5 h. Compound 18a (986 mg, 823 µL, 8.0 mmol, 2.0 equiv.) was then added dropwise, and stirring was continued for 14 h. After addition of saturated aqueous NH_4Cl (pH = 7) the mixture was extracted with EtOAc ($3 \times 100 \text{ mL}$). The combined organic layers were washed with H_2O (3 × 100 mL) and brine (3 × 100 mL), dried (MgSO₄) and concentrated in vacuo. Purification of the residue by column chromatography (SiO2; PE/EtOAc, 8:1) afforded 19a as a yellow solid (1.76 g, 3.06 mmol, 77%). ¹H NMR (300 MHz, CDCl₃): $\delta = 1.22-1.35$, 1.49–1.60 [2 × m, total 9 H, C(CH₃)₃], 3.69– 3.83, 4.17–4.33 ($2 \times m$, total 1 H, 1'-H_a), 4.42–4.61 (m, 1 H, 1'- H_b), 5.10–5.34 (m, 2 H, OC H_2 Ph), 5.90–3.12 (m, 1 H, 2'-H), 6.74– 6.92 (m, 1 H, 3-H), 7.35–7.69 (m, 5 H, 5× Ph-H), 7.58–7.69 (m, 1 H, 6-H), 8.32-8.44 (m, 1 H, 5-H), 8.57-8.64 (m, 1 H, 8-H) ppm. ¹³C NMR (150 MHz, CDCl₃): $\delta = 28.12$, 28.14 [C(CH₃)₃], 46.11, 49.19 (C-1'), 71.01, 71.10 (OCH₂Ph), 81.01, 81.07 [C(CH₃)₃], 94.57, 94.66 (C-1), 110.41, 110.82 (C-3), 112.19, 112.26 (C-7), 118.31, 118.35 (CN), 120.57, 121.72 (C-3'), 124.03, 124.07 (C-5), 126.7, 126.8, 127.0, 127.1, 127.2, 128.2, 128.5, 128.6, 129.5, 134.8, 134.9, 135.6, 135.7, 144.8, 145.1 (C-4a, C-8a, C-2, C-2', 6 × Ph-C), 138.3, 138.4 (C-8), 152.9, 153.1, 155.2 (C-4, C=O) ppm. IR (KBr): \tilde{v} = 2976, 2228, 1703, 1593, 1503, 1410, 1368, 1329, 1257, 1163 cm⁻¹.

UV (CH₃CN): λ_{max} (lg ε) = 221.0 (4.645), 256.0 (4.509), 320.5 (3.828), 354.4 (3.638) nm. ESI-HRMS: m/z = 597.04123 [M + Na] $^+$; C₂₆H₂₄ClIN₂O₃ requires 597.04187.

(E/Z)-2-Amino-4-(benzyloxy)-1-bromo-N-(tert-butoxycarbonyl)-N-(3-chlorobut-2-enyl)-6-cyanonaphthalene (19b): NaH (863 mg, 60% dispersion in mineral oil, 21.6 mmol, 2.5 equiv.) was added at room temperature to a solution of 17b (6.52 g, 14.4 mmol, 1.0 equiv.) in anhydrous DMF (150 mL) and the mixture was stirred for 3 h. Compound 18b (3.59 g, 3.16 mL, 28.8 mmol, 2.0 equiv.) was then added dropwise, and stirring was continued for 4 h. After addition of saturated aqueous NH₄Cl (200 mL) and H₂O (100 mL), the mixture was extracted with EtOAc (3× 200 mL). The combined organic layers were washed with H₂O (3× 200 mL) and brine (3× 200 mL), dried (MgSO₄) and concentrated in vacuo. The residue was washed with n-pentane (150 mL) and recrystallised from EtOAc. Workup of the mother liquor by column chromatography (SiO₂; PE/EtOAc, 10:1) afforded another batch of **19b** as a white solid (7.48 g, 13.8 mmol, total 96%). ¹H NMR (300 MHz, CDCl₃): $\delta = 1.26-1.36$, 1.52-1.62 [2 × m, total 9 H, C(CH₃)₃], 1.81-1.87, 1.99-2.10 (2 × m, total 3 H, 4'-H₃), 3.90-4.03, 4.23-4.38 (2 × m, total 1 H, 1'-H_a), 4.39–4.55 (m, 1 H, 1'-H_b), 5.13–5.37 (m, 2 H, OCH_2Ph), 5.61–5.83 (m, 1 H, 2'-H), 6.74–6.96 (m, 1 H, 3-H), 7.37– $7.57 \text{ (m, 5 H, 5} \times \text{Ph-H)}, 7.69 - 7.81 \text{ (m, 1 H, 7-H)}, 8.33 - 8.41 \text{ (m, 1 H, 7-H)}$ H, 8-H), 8.65-8.76 (m, 1 H, 5-H) ppm. ¹³C NMR (75 MHz, CDCl₃): δ = 20.84, 26.15 (C-4'), 28.11, 28.38 [C(*C*H₃)₃], 46.19, 47.48 (C-1'), 70.83, 70.90 (OCH₂Ph), 80.94 [C(CH₃)₃], 108.8, 109.4, 109.6, 109.8 (C-3, C-6), 114.5, 118.8, 118.9, (C-4a, CN), 121.2, 122.5 (C-2'), 124.9 (C-1), 127.4, 127.7, 128.5, 128.7, 128.8, 128.9 $(C-5, C-7, C-8, 2 \times Ph-C_o, 2 \times Ph-C_m, Ph-C_p), 133.3, 134.2, 135.4,$ 135.5, 142.1, 142.2 (C-2, C-8a, C-3', Ph-C_i), 153.6, 154.1, 154.6 (C-4, C=O) ppm. IR (KBr): $\tilde{v} = 3083$, 2967, 2223 (CN), 1702 (C=O), 1617, 1592 cm⁻¹. UV (CH₃CN): λ_{max} (lg ε) = 218.0 (4.655), 256.5 (4.587), 318.0 (3.758), 335.5 (3.762), 350.5 (3.840) nm. ESI-HRMS: $m/z = 541.0888 [M + H]^+$; $C_{27}H_{26}CIIN_2O_3$ requires 541.0888.

(2R)-2-Amino-4-(benzyloxy)-1-bromo-N-(tert-butoxycarbonyl)-7-cyano-N-(2,3-epoxypropyl)naphthalene [(2R)-21a]: NaH (173 mg, 60% dispersion in mineral oil, 4.4 mmol, 4.0 equiv.) was added to a solution of 17a (500 mg, 1.10 mmol, 1.0 equiv.) in anhydrous DMF (5 mL), and the mixture was stirred at room temperature for 30 min. The nosylate (2R)-20a (456 mg, 1.76 mmol, 1.6 equiv.) was added, and stirring was continued for 14 h. The reaction was quenched by addition of saturated aqueous NaHCO₃ (50 mL) and brine (50 mL). The reaction mixture was extracted with EtOAc (3 \times 50 mL), and the combined organic layers were dried (MgSO₄) and concentrated in vacuo. Purification of the residue by column chromatography (SiO₂; PE/EtOAc, 5:1) afforded (2R)-21a as a bright yellow solid (495 mg, 972 μmol, 88%). ¹H NMR (300 MHz, $C_2D_2Cl_4$): $\delta = 1.20-1.72$ [m, 9-H, $C(CH_3)_3$], 2.38-2.50 (m, 1 H, 4'-H_a) 2.67–2.81 (m, 1 H, 4'-H_b), 3.08–3.15 (m, 0.5 H, 2'-H_a), 3.15– 3.28 (m, 0.5 H, 3'-H), 3.30–3.40 (m, 0.5 H, 2'-H_a), 3.51–3.73 (m, 0.5 H, 3'-H), 3.88-4.21 (m, 1 H, 2'-H_b), 5.22-5.35 (m, 2 H, 1) OCH_2Ph), 6.93–7.13 (m, 1 H, 3-H), 7.34–7.54 (m, 5 H, 5× Ph-H), 7.64-7.72 (m, 1 H, 6-H), 8.40-8.49 (m, 1 H, 5-H), 8.64-8.69 (m, 1 H, 8-H) ppm. IR (KBr): $\tilde{v} = 2977$, 2228, 1703, 1596, 1503, 1370, 1257, 1153 cm⁻¹. UV (CH₃CN): λ_{max} (lg ε) = 215.5 (4.618), 256.5 (4.596), 316.0 (3.815), 352.0 (3.580) nm. ESI-HRMS: m/z = $509.10705 [M + H]^{+}$; $C_{26}H_{25}BrN_2O_4$ requires 509.10759.

(+)-(2*R*,3*R*)-2-Amino-4-(benzyloxy)-1-bromo-*N*-(*tert*-butoxycarbonyl)-6-cyano-*N*-(2,3-epoxybutyl)naphthalene [(+)-(2*R*,3*R*)-21b]: NaH (397 mg, 60% dispersion in mineral oil, 16.6 mmol, 4.0 equiv.) was added to a solution of 17c (1.88 g, 4.14 mmol, 1.0 equiv.) in anhydrous DMF (30 mL), and the mixture was stirred

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at 60 °C for 60 min. After the system had cooled to room temperature, the nosylate (2R,3R)-**20b** (1.70 g, 6.21 mmol, 1.5 equiv.) was added, and stirring was continued for 2 h. The reaction was quenched by addition of saturated aqueous NH₄Cl (100 mL). The reaction mixture was extracted with Et₂O (3× 100 mL), and the combined organic layers were dried (Na₂SO₄) and concentrated in vacuo. Purification of the residue by column chromatography (SiO₂; PE/EtOAc, 9:1) afforded (+)-(2R,3R)-21b as a white solid (1.93 g, 3.69 mmol, 89%). ¹H NMR $(300 \text{ MHz}, \text{CDCl}_3)$: $\delta = 1.25$ – 1.41, 1.59–1.68 [2 \times m, total 12 H, C(CH₃)₃, 4'-H₃], 2.60–2.62, 2.76-2.91, 3.04-3.22, 3.27-3.36, 3.52-3.61, 3.71-4.21, 4.30-4.35 $(7 \times m, \text{ total 4 H}, 1'-H_2, 2'-H, 3'-H), 5.20-5.32 (m, 2 H, OCH_2Ph),$ 6.93-7.10 (m, 1 H, 3-H), 7.37-7.54 (m, 5 H, 5 × Ph-H), 7.69-7.76 (m, 1 H, 7-H), 8.18-8.21, 8.33-8.38 (2 × m, total 1 H, 8-H), 8.66-8.72 (m, 1 H, 5-H) ppm. ¹³C NMR (125 MHz, CDCl₃): $\delta = 17.2$ (2 signals, C-4'), 28.0, 28.1, 28.4 (2 signals, C(CH₃)₃], 50.5, 52.1, 52.4, 53.0, 53.4, 53.6, 53.7, 54.2, 56.5, 56.8, 57.1, 57.2 (C-1', C-2', C-3'), 68.9, 70.9 (OCH₂Ph), 81.0, 81.1, 81.7 (2 signals, C(CH₃)₃], 109.0, 109.1, 109.3, 109.6, 109.7 (C-3, C-6), 114.0, 114.5, 114.6, 118.9, (2 signals, C-4a, CN), 125.0, 125.1, 125.9 (C-1), 127.5, 127.6, 127.8, 127.9, 128.5 (2 signals), 128.7 (2 signals), 128.8 (2 signals), 128.9 (2 signals, C-5, C-7, C-8, $2 \times \text{Ph-C}_{o}$, $2 \times \text{Ph-C}_{m}$, Ph-C_p), 134.1 (2 signals), 134.3, 134.4, 135.4, 135.5, 142.1, 142.2, 142.6 (C-2, C-8a, Ph-C_i), 153.5, 153.6, 153.7, 154.2 (2 signals), 154.7, 163.5 (C-4,C=O) ppm. IR (film): $\tilde{v} = 2977$, 2228, 1706, 1620, 1591, 1455, 1411, 1369, 1342, 1254, 1151, 1095, 975, 910, 872, 738, 697 cm⁻¹. UV (CH₃CN): λ_{max} (lg ε) = 218.0 (4.630), 256.5 (4.557), 317.0 (3.812), 334.5 (3.765), 350.5 (3.814) nm. ESI-HRMS: m/z = $545.1046 [M + Na]^+$; $C_{27}H_{27}BrN_2O_4$ requires 545.1046.

rac-(1S)-5-(Benzyloxy)-3-(tert-butoxycarbonyl)-1-(chloromethyl)-8cyano-1,2-dihydro-3H-benz[e]indole (rac-22a): A solution of 19a (1.42 g, 2.47 mmol, 1.0 equiv.) in anhydrous toluene (60 mL) was flushed with argon for 10 min. Tris(trimethylsilyl)silane (836 µL, 2.72 mmol, 1.1 equiv.) and AIBN (102 mg, 618 µmol, 0.25 equiv.) were added at room temperature, and the solution was stirred at 80 °C with use of a preheated oil bath for 4.5 h. After addition of SiO₂ (3 g), the mixture was concentrated in vacuo, and the residue was purified by column chromatography (n-hexane/DCM, 9:1) to afford rac-22a as a white solid (940 mg, 2.10 mmol, 85%). ¹H NMR (300 MHz, $C_2D_2Cl_4$): $\delta = 1.64$ [s, 9 H, $C(CH_3)_3$], 3.59 (dd, J = 9.1, 11.2 Hz, 1 H, 2-H_a), 3.89 (dd, J = 3.51, 11.2 Hz, 1 H, 2- H_b), 4.00 (tt, J = 3.2, 8.8 Hz, 1 H, 1-H), 4.21 (dd, J = 8.8, 11.7 Hz, 1 H, 10-H_a), 4.30 (dd, J = 3.2, 11.8 Hz, 1 H, 10-H_b), 5.32 (s, 2 H, OCH_2Ph), 7.36–7.57 (m, 6 H, 7-H, 5× Ph-H), 7.93 (s, 1 H, 4-H), 8.02 (dd, J = 0.7, 1.5 Hz, 1 H, 9-H), 8.38 (dd, J = 0.5, 8.7 Hz, 1H, 6-H) ppm. ¹³C NMR (100 MHz, $C_2D_2Cl_4$): $\delta = 28.26$ [C(CH₃) ₃], 41.01 (C-1), 46.37 (C-2), 53.15 (C-10), 70.84 (O*C*H₂Ph), 81.57 [C(CH₃)₃], 99.71 (C-4), 111.1 (C-8), 114.9 (C-9b), 118.9 (CN), 123.2 (C-7), 123.6 (C-5a), 124.9 (C-6), 127.4 (C-9), 127.4 (2 \times Ph-C_o), 128.0 (Ph- C_p), 128.4 (2 × Ph- C_m), 129.4 (C-9a), 136.1 (Ph- C_i), 143.4 (C-3a), 151.9 (C=O), 155.9 (C-5) ppm. IR (KBr): $\tilde{v} = 2977$ 2230, 1693, 1593, 1455, 1409, 1374, 1336, 1257, 1146 cm⁻¹. UV (CH₃CN): λ_{max} (lg ε) = 210.0 (4.391), 230.0 (4.469), 268.0 (4.786), 311.5 (3.906), 322.5 (3.977), 381.0 (3.475) nm. ESI-HRMS: m/z = $471.14459 [M + Na]^+$; $C_{26}H_{25}CIN_2O_3$ requires 471.14459.

Chromatographic Resolution of *rac*-22a: A solution of *rac*-22a (500 mg, 1.1 mmol) in a mixture of *n*-hexane and CH₂Cl₂ (1:1, 10 mL) was separated (injection volume: 0.15 mL) by semipreparative HPLC (Chiralpak IA; 250×20 mm; particle size: 5 µm; *n*-hexane/CH₂Cl₂, 85:15; flow: 18 mL min⁻¹; UV detector: λ = 269 nm) to provide (+)-(1*R*)-22a (t_R = 12.5 min) and (-)-(1*S*)-22a (t_R = 10.8 min). The optical purity was determined by analytical HPLC (Chiralcel OD; 250×4.6 mm; particle size: 10 µm; *n*-hex-

ane/iPrOH, 97:3; flow: 0.8 mL min⁻¹): (+)-(1*R*)-22a: 99.7% ee (t_R = 8.4 min), [a]_D²⁰ = +9.5 (c = 0.5 in CHCl₃); (-)-(1*S*)-22a: 99.2% ee (t_R = 6.4 min), [a]_D²⁰ = -11.0 (c = 0.5 in CHCl₃).

rac-(1S)-5-(Benzyloxy)-3-(tert-butoxycarbonyl)-1-[(1R)-1-chloroethyl]-7-cyano-1,2-dihydro-3*H*-benz[*e*]indole (*rac*-22b) and *rac*-(1*S*)-5-(Benzyloxy)-3-(tert-butoxycarbonyl)-1-[(1S)-1-chloroethyl]-7-cyano-1,2-dihydro-3*H*-benz[e]indole (rac-22c): A solution of 19b (3.20 g, 5.91 mmol, 1.0 equiv.) in anhydrous toluene (150 mL) was flushed with argon for 10 min. Tris(trimethylsilyl)silane (2.00 mL, 6.48 mmol, 1.1 equiv.) and AIBN (242 mg, 1.47 mmol, 0.25 equiv.) were added at room temperature, and the solution was stirred at 80 °C with use of a preheated oil bath for 4.5 h. After the system had cooled to room temperature, the brown solid was washed with *n*-pentane. The resulting crude solid was purified by column chromatography (PE/EtOAc, $30:1 \rightarrow 10:1$) to afford the syn diastereomer rac-22c as a white solid (560 mg, 1.21 mmol, 20%), a mixture of the syn diastereomer rac-22c together with the dehalogenised cyclisation product as a yellow solid (380 mg) and the desired anti diastereomer rac-22b as a white solid (1.30 g, 2.81 mmol, 48%).

rac-22c: ¹H NMR (300 MHz, $C_2D_2Cl_4$, 100 °C): $\delta = 1.60-1.67$ [m, 12 H, 11-H₃, C(CH₃)₃], 3.87 (m_c, 1 H, 1-H), 4.13 (dd, J = 11.8, 9.5 Hz, 1 H, 2-H_a), 4.37 (dd, J = 11.8, 2.8 Hz, 1 H, 2-H_b), 4.55 $(dq, J = 6.8, 3.7 \text{ Hz}, 1 \text{ H}, 10\text{-H}), 5.33 (m_c, 2 \text{ H}, OCH_2Ph), 7.37$ 7.57 (m, 5 H, 5 \times Ph-H), 7.60 (dd, J = 8.7, 1.6 Hz, 1 H, 8-H), 7.72 (d, J = 8.7 Hz, 1 H, 9 -H), 7.87 (br. s, 1 H, 4-H), 8.66 (d, J = 1.6 Hz,1 H, 6-H) ppm. ¹³C NMR (75 MHz, $C_2D_2Cl_4$, 100 °C): $\delta = 23.04$ (C-11), 28.24 [C(CH₃)₃], 45.54 (C-1), 50.95 (C-2), 59.68 (C-10), 70.79 (OCH₂Ph), 81.56 [C(CH₃)₃], 98.24 (C-4), 105.5 (C-7), 115.5 (C-9b), 119.4, 121.4 (C-5a, CN), 123.0 (C-9), 127.4, 127.6, 128.1, 128.5 (C-8, $2 \times \text{Ph-C}_o$, $2 \times \text{Ph-C}_m$, Ph-C_p), 129.8 (C-6), 131.7 (C-9a), 135.9 (Ph-C_i), 145.1 (C-3a), 151.7 (C=O), 156.1 (C-5) ppm. IR (KBr): $\tilde{v} = 2976$, 2223 (CN), 1705 (C=O), 1622, 1577, 1478, 1456 cm⁻¹. UV (CH₃CN): λ_{max} (lg ε) = 214.0 (4.452), 270.5 (4.829), 353.0 (4.198) nm. ESI-HRMS: $m/z = 463.1783 \, [M + H]^+$; C₂₆H₂₅ClN₂O₃ requires 463.1783.

Chromatographic Resolution of *rac*-22b: A solution of *rac*-22b (1.0 g, 2.2 mmol) in a mixture of *n*-heptane/CH₂Cl₂ (1:1, 25 mL) was separated (injection volume: 1.0 mL) by semipreparative HPLC (Chiralpak IA; 250×20 mm; particle size: 5 µm; *n*-heptane/ CH₂Cl₂, 80:20; flow: 18 mL min⁻¹; UV detector: λ = 250 nm) to provide (+)-(1*S*,10*R*)-22b (t_R = 8.2 min) and (-)-(1*R*,10*S*)-22b (t_R = 10.8 min). The optical purity was determined by analytical HPLC (Chiralcel OD; 250×4.6 mm; particle size: 10 µm; *n*-hexane/*i*P-rOH, 98:2; flow: 0.8 mL min⁻¹): (+)-(1*S*,10*R*)-22b: 99.9% *ee* (t_R = 37.6 min), [a]²⁰_D = +24.6 (c = 0.5 in CHCl₃); (-)-(1*R*,10*S*)-22b: 98.7% *ee* (t_R = 33.9 min), [a]²⁰_D = -24.4 (c = 0.5 in CHCl₃).

(1.S)-5-(Benzyloxy)-3-(tert-butoxycarbonyl)-8-cyano-1-(hydroxymethyl)-1,2-dihydro-3*H*-benz[e]indole [(1.S)-23a]: A solution of *n*-butyllithium in *n*-hexane (2.5 m, 245 μL, 612 μmol, 3.0 equiv.) was added slowly at -90 °C to a stirred solution of (2*R*)-21a (104.0 mg, 20.4 mmol, 1.0 equiv.) in anhydrous THF (10 mL), and stirring was continued at -78 °C for 1 h. The reaction was quenched by addition of saturated aqueous NH₄Cl (50 mL), and the mixture was extracted with EtOAc (2× 100 mL). The combined organic layers were washed with brine (300 mL), dried (MgSO₄) and concentrated in vacuo. Purification of the crude solid by column chromatography (SiO₂; PE/EtOAc, 4:1) afforded (1.S)-23a as a bright brown solid (10.1 mg, 2.35 mmol, 12%). ¹H NMR (300 MHz, CDCl₃): δ = 1.58 [s, 9 H, C(CH₃)₃], 3.66–4.26 (m, 5 H, 1-H, 2-H, 10-H), 5.23 (s, 2 H, OCH₂Ph), 7.31–7.55 (m, 6 H, 7-H, 5× Ph-H), 7.94–8.05 (m, 1 H, 4-H), 8.05–8.08 (m, 1 H, 9-H), 8.26–8.35 (m, 1 H, 6-



H) ppm. ¹³C NMR (150 MHz, CDCl₃): δ = 28.38 [C(CH₃)₃], 41.13 (C-1), 52.48 (C-2), 64.79 (C-10), 70.54 (OCH₂Ph), 81.60 [C(CH₃)₃], 99.10 (C-4), 110.6 (C-8), 115.0 (C-9b), 119.4 (CN), 123.2 (C-7), 123.5 (C-5a), 124.8 (C-6), 127.4 (C-9), 127.7 (2 × Ph-C_o), 128.2 (Ph-C_p), 128.6 (2 × Ph-C_m), 129.7 (C-9a), 136.2 (Ph-C_i), 143.7 (C-3a), 152.5 (C=O), 155.4 (C-5) ppm. IR (KBr): \hat{v} = 2974, 2227, 1701, 1623, 1291, 1455, 1369, 1257, 1143, 1092, 1031 cm⁻¹. UV (CH₃CN): λ_{max} (Ig ε) = 209.0 (4.393), 229.0 (4.435), 267.5 (4.683), 313.0 (3.848), 323.5 (3.878), 375.0 (3.405) nm. ESI-HRMS: m/z = 431.19653 [M + H]⁺; C₂₆H₂₆N₂O₄ requires 431.19653.

(+)-(1S)-5-(Benzyloxy)-3-(tert-butoxycarbonyl)-7-cyano-1-[(1R)-1hydroxyethyl]-1,2-dihydro-3H-benz[e]indole [(+)-(1S,10R)-23b]: Freshly fused ZnCl₂ (544 mg, 3.99 mmol, 1.5 equiv.) was suspended in anhydrous THF (50 mL), and the mixture was cooled to 0 °C. A solution of methyllithium in Et₂O (1.6 M, 9.98 mL, 16.0 mmol, 6.0 equiv.) was slowly added to the stirred mixture, and stirring was continued at 0 °C for 30 min. The mixture was cooled to -78 °C, and TMS-NCS (563 µL, 524 mg, 3.99 mmol, 1.5 equiv.) was added dropwise. The mixture was warmed to 0 °C and stirred for 30 min. After the mixture had again been cooled to -78 °C, a solution of (+)-(2'R,3'R)-**21b** (1.39 g, 2.66 mmol, 1.0 equiv.) in anhydrous THF (25 mL) was added dropwise, and the resulting mixture was stirred at -78 °C for 30 min. After warming to 0 °C, the mixture was stirred for 1 h, and, after warming to room temperature, it was stirred for 2.5 h. Saturated aqueous NH₄Cl (200 mL) was slowly added, and the mixture was extracted with CH_2Cl_2 (4× 200 mL). The combined organic layers were dried (Na₂SO₄) and concentrated in vacuo. Purification of the residue by column chromatography (SiO₂; PE/EtOAc, 5:2) afforded (+)-(1S,10R)-23b as a white solid (840 mg, 1.89 mmol, 71%, 98.8% ee). ¹H NMR (300 MHz, $C_2D_2Cl_4$, 100 °C): $\delta = 1.27$ (d, J = 6.4 Hz, 3 H, 10-CH₃), 1.63 [br. s, 9 H, C(CH₃)₃], 3.71 (dt, J = 9.9, 3.1 Hz, 1 H, 1-H), 4.09 (dd, J= 11.6, 9.9 Hz, 1 H, 2-H_a), 4.26 (dd, J = 11.6, 3.4 Hz, 1 H, 2-H_b), 4.33 (m_c, 1 H, 10-H), 5.32 (s, 2 H, OCH₂Ph), 7.38–7.54 (m, 5 H, $5 \times \text{Ph-H}$), 7.58 (dd, J = 8.8, 1.7 Hz, 1 H, 8-H), 7.83 (d, J = 8.8 Hz, 1 H, 9-H), 7.89 (br. s, 1 H, 4-H), 8.64 (d, J = 1.6 Hz, 1 H, 6-H) ppm. ¹³C NMR (150 MHz, $C_2D_2Cl_4$, 100 °C): $\delta = 20.2$ (C-11), 28.3 [C(CH₃)₃], 45.0 (C-1), 49.9 (C-2), 68.7 (C-10), 70.8 (OCH₂Ph), 81.4 [C(CH₃)₃], 98.3 (C-4), 105.5 (C-7), 115.7 (C-9b), 119.5, 121.4 (C-5a, CN), 123.7 (C-9), 127.4 (2 signals), 128.1, 128.5 (C-8, 5× Ph-CH), 129.7 (C-6), 132.0 (C-9a), 136.0 (Ph-C_i), 145.2 (C-3a), 151.8 (C=O), 155.9 (C-5) ppm. IR (KBr): $\tilde{v} = 2973$, 2222, 1702, 1620, 1577, 1479, 1455, 1413, 1373, 1325, 1255, 1144, 909, 822, 736, 696 cm⁻¹. UV (CH₃CN): λ_{max} (lg ε) = 215.0 (4.462), 271.5 (4.835), 355.5 (4.211) nm. ESI-HRMS: m/z = 445.2121 [M + H]⁺; C₂₇H₂₈N₂O₄ requires 445.2122.

(+)-(1*S*)-5-(Benzyloxy)-3-(*tert*-butoxycarbonyl)-1-[(1*R*)-1-chloroethyl]-7-cyano-1,2-dihydro-3*H*-benz[*e*]indole [(+)-(1*S*,10*R*)-22b]: SOCl₂ (206 μL, 337 mg, 2.83 mmol, 10.0 equiv.) and pyridine (572 μL, 560 mg, 7.09 mmol, 25.0 equiv.) were added at 0 °C to a solution of (+)-(1*S*,10*R*)-23b (126 mg, 283 μmol, 1.0 equiv.) in anhydrous CH₂Cl₂ (6 mL). The reaction mixture was stirred at room temperature for 4 h. After concentration, the crude product was purified by column chromatography (PE/EtOAc, 5:1) to afford (+)-(1*S*,10*R*)-22b as a white solid (57.8 mg, 164 μmol, 58 %, 99.0 % *ee*).

(-)-(1*S*)-3-(*tert*-Butoxycarbonyl)-1-(chloromethyl)-8-cyano-5-hydroxy-1,2-dihydro-3*H*-benz[*e*]indole [(-)-(1*S*)-24a]: Pd/C (10%, 140 mg, 129 μ mol, 0.1 equiv. based on Pd) was added to a solution of (-)-(1*S*)-22a (250 mg, 557 μ mol, 1.0 equiv.) in anhydrous EtOAc (17 mL). The mixture was flushed with argon and H₂, and the mixture was stirred at room temperature under H₂ for 5 h. The reaction mixture was filtered through Celite, the filter cake was washed with

EtOAc (800 mL), and the filtrate was concentrated in vacuo. Purification of the residue by column chromatography (SiO₂; PE/EtOAc, 5:1) afforded (-)-(1S)-24a as a yellow solid (179 mg, 499 μ mol, 90%). ¹H NMR (300 MHz, [D₆]DMSO): $\delta = 1.54$ [s, 9 H, C(CH₃) ₃], 3.77–4.31 (m, 5 H, 1-H, 2-H₂, 10-H₂), 7.50 (dd, J = 1.5, 8.7 Hz, 1 H, 7-H), 7.72 (br. s, 1 H, 4-H), 8.18 (d, J = 8.7 Hz, 1 H, 6-H), 8.41 (dd, J = 0.5, 1.5 Hz, 1 H, 9-H), 10.88 (br. s, 1 H, OH) ppm. ¹³C NMR (75 MHz, [D₆]DMSO): $\delta = 27.98$ [C(CH₃)₃], 47.86 (C-1), 52.50 (C-2), 61.92 (C-10), 80.8 [C(CH₃)₃], 101.0 (C-4), 109.7 (C-8), 114.1 (C-9b), 119.2 (CN), 122.0 (C-5a), 122.5 (C-7), 124.5 (C-6), 128.8 (C-9), 129.2 (C-9a), 143.2 (C-3a), 151.5 (C=O), 154.6 (C-5) ppm. IR (KBr): $\tilde{v} = 3350$, 2230, 1676, 1626, 1595, 1513, 1422, 1369, 1223, 1144 cm⁻¹. UV (CH₃CN): λ_{max} (lg ε) = 210.5 (4.221), 229.5 (4.369), 269.0 (4.787), 312.5 (3.826), 323.5 (3.861), 381.0 (3.428) nm. ESI-HRMS: m/z = 381.09764 [M + Na]⁺; C₁₉H₁₉ClN₂O₃ requires 381.09764.

Chromatographic Resolution of (-)-(1*S*)-24a: A solution of (-)-(1*S*)-24a (250 mg, 697 µmol) in a mixture of *n*-hexane/*i*PrOH (1:1, 10 mL) was separated consecutively (injection volume: 0.5 mL) by semipreparative HPLC (Chiralpak IA; 250×20 mm; particle size: 5 µm; *n*-hexane/*i*PrOH, 98:2; flow: 18 mL min⁻¹; UV detector: $\lambda = 250$ nm) to provide (-)-(1*S*)-24a ($t_R = 19.4$ min), $[a]_D^{20} = -20.0$ (c = 0.5 in CHCl₃).

(+)-(1S)-3-(tert-Butoxycarbonyl)-1-[(1R)-1-chloroethyl]-7-cyano-5hydroxy-1,2-dihydro-3*H*-benz[*e*]indole [(+)-(1*S*,10*R*)-24b]: A solution of (+)-(1S,10R)-22b (130 mg, 281 µmol, 1.0 equiv.) in anhydrous THF (9 mL) was warmed to 40 °C and Pd/C (10%, 30 mg, 28 µmol, 0.1 equiv. based on Pd) was added. A solution of NH₄CHO₂ in H₂O (25%, 0.30 mL) was added dropwise, and the mixture was stirred at 40 °C for 25 min. The mixture was filtered through Celite, the filter cake was washed with acetone (300 mL), and the filtrate was concentrated in vacuo. Purification of the residue by column chromatography (SiO₂; PE/EtOAc, $10:1 \rightarrow 5:1 \rightarrow 3:1$) afforded (+)-(1S,10R)-24b as a white solid (92 mg, 247 µmol, 88%). $[a]_{D}^{20} = +22.8$ (c = 0.5 in CHCl₃). ¹H NMR (300 MHz, $[D_6]DMSO$): $\delta = 1.54$ [s, 9 H, C(CH₃)₃], 1.58 (d, J = 6.8 Hz, 3 H, 11-H₃), 3.98– 4.16 (m, 3 H, 1-H, 2-H₂), 4.71 (m_c, 1 H, 10-H), 7.65 (m_c, 2 H, 4-H, 8-H), 7.93 (d, J = 8.8 Hz, 1 H, 9-H), 8.46 (d, J = 1.2 Hz, 1 H, 6-H), 10.89 (br. s, 1 H, OH) ppm. 13 C NMR (75 MHz, [D₆]DMSO): δ = 22.92 (C-11), 27.96 [C(CH₃)₃], 44.07 (C-1), 49.46 (C-2), 61.24 (C-10), 80.93 [C(CH₃)₃], 99.61 (C-4), 103.8 (C-7), 114.7 (C-9b), 119.6, 119.9 (C-5a, CN), 123.9 (C-9), 127.2 (C-8), 129.5 (C-6), 131.3 (C-9a), 144.7 (C-3a), 151.3 (C=O), 155.1 (C-5) ppm. UV (CH₃CN): $\lambda_{\text{max}} (\lg \varepsilon) = 214.5 (4.347), 271.5 (4.820), 355.5 (4.148) \text{ nm. IR}$ (KBr): $\tilde{v} = 3361$ (C=O), 2978, 2221 (CN), 1687 (C=O), 1623, 1577 cm⁻¹. ESI-HRMS: $m/z = 373.1314 \text{ [M + H]}^+$; $C_{20}H_{21}ClN_2O_3$ requires 373.1314.

(1.5)-1-(Chloromethyl)-8-cyano-3-({5-[2-(dimethylamino)ethoxy]indol-2-yl}carbonyl)-1,2-dihydro-3H-benz[e|indol-5-yl 2,3,4,6-Tetra-O-acetyl- β -D-galactopyranoside [(+)-(1.5)-27a]: A mixture of the phenol (-)-(1.5)-24a (283 mg, 663 μmol, 1.00 equiv.) in dry CH₂Cl₂ (20 mL) and freshly activated molecular sieves (4 Å, 2.00 g) was stirred at room temperature for 30 min. After addition of the trichloroacetimidate 25 (427 mg, 870 μmol, 1.3 equiv.) and cooling to -10 °C, the promoter BF₃·OEt₂ (50 μL) in dry CH₂Cl₂ (4 mL) was added dropwise, and the mixture was stirred for 3.5 h. Additional BF₃·OEt₂ (250 μL) in CH₂Cl₂ (4 mL) was then added, and the mixture was allowed to warm to room temperature and stirred for 6 h. The reaction mixture was transferred by cannula into a new flask for separation of the molecular sieves, which were washed with CH₂Cl₂ (30 mL). The obtained organic solution was concentrated in vacuo, and the residue was dissolved in DMF (38 mL) and co-

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oled to 0 °C. EDC·HCl (450 mg, 2.36 mmol, 3.5 equiv.) and DMAI·HCl (26, 336 mg, 1.35 mmol, 2.0 equiv.) were added, and the reaction mixture was stirred at room temperature for 18 h. After dilution with EtOAc (70 mL), H₂O (70 mL) and saturated aqueous NaHCO₃ (70 mL) were added, and the aqueous phase was extracted with EtOAc (4× 150 mL). The combined organic layers were washed with brine (4 × 200 mL), dried (MgSO₄) and concentrated in vacuo. Purification of the residue by column chromatography (SiO₂; CH₂Cl₂/MeOH, 6:1) afforded (+)-(1S)-27a as a white solid (340 mg, 416 μ mol, 53%). $[a]_D^{20} = +6.0$ (c = 0.3 in CHCl₃). ¹H NMR (600 MHz, CDCl₃): $\delta = 2.01$ [s, 6 H, 2× C(O)CH₃], 2.03 [s, 3 H, C(O)CH₃], 2.16 [s, 3 H, C(O)CH₃], 2.37 (s, 6 H, NMe₂), 2.79 $(t, J = 5.7 \text{ Hz}, 2 \text{ H}, 2'' - \text{H}_2), 3.55 \text{ (dd}, J = 11.5, 10.0 \text{ Hz}, 1 \text{ H}, 10-10.0 \text{ Hz}, 1 \text{ Hz}$ H_a), 3.78–3.85 (m, 1 H, 1-H), 3.90 (dd, J = 11.5, 3.2 Hz, 1 H, 10-H_b), 4.05–4.23 (m, 5 H, 1"-H₂, 5""-H, 6""-H₂), 4.69–4.75 (m, 1 H, $2-H_a$), 4.84 (dd, J = 10.6, 2.0 Hz, 1 H, $2-H_b$), 4.99 (dd, J = 10.5, 3.2 Hz, 1 H, $3^{\prime\prime\prime}$ -H), 5.27 (d, J = 7.9 Hz, 1 H, $1^{\prime\prime\prime}$ -H), 5.38 (d, J =3.3 Hz, 1 H, $4^{\prime\prime\prime}$ -H), 5.63 (dd, J = 10.5, 7.9 Hz, 1 H, $2^{\prime\prime\prime}$ -H), 7.03– 7.06 (m, 2 H, 3'-H, 6'-H), 7.14 (d, J = 2.2 Hz, 1 H, 4'-H), 7.37 (d, J = 8.9 Hz, 1 H, 7'-H, 7.53 (dd, J = 8.7, 1.5 Hz, 1 H, 7-H, 8.08(d, J = 0.8 Hz, 1 H, 9 -H), 8.21 (d, J = 8.7 Hz, 1 H, 6 -H), 8.51 (s, 1)1 H, 4-H), 10.05 (s, 1 H, NH) ppm. ¹³C NMR (125 MHz, CDCl₃): δ = 20.63, 20.69, 20.72, 20.84 [4× C(O)*C*H₃], 45.84 (NMe₂), 45.96 (C-10), 54.99 (C-2), 58.35 (C-2"), 61.16 (C-6""), 66.33 (C-1"), 66.78 (C-4'''), 68.43 (C-2'''), 70.51 (C-3'''), 71.20 (C-1), 99.02 (C-1'''), 103.52 (C-4'), 103.9 (C-4), 106.4 (C-6'*), 111.6 (C-5a), 112.7 (C-7'), 117.8 (C-3'*), 118.6 (C-8), 118.8, 124.4 (C-5a, C-9b), 124.8 (C-6), 125.0 (C-7), 128.0 (C-9), 128.2, 128.7, 130.0, 131.5 (CN, C-2', C-3a', C-7a'), 143.7 (C-3a), 153.3 (C-5), 153.9 (C-5'), 160.8 (C=O), 169.5, 169.7, 170.0, 170.3 [$4 \times C(O)CH_3$] ppm. IR (KBr): $\tilde{v} = 3406 \text{ (NH)}, 2942, 2229 \text{ (CN)}, 1754 \text{ (C=O)}, 1626, 1521, 1457,$ 1396, 1226 cm⁻¹. UV (CH₃CN): λ_{max} (lg ε) = 212.5 (4.712), 261.5 (4.431), 302.0 (4.546), 399.0 (4.519) nm. ESI-HRMS: m/z =819.2651 [M + H]⁺; $C_{41}H_{43}ClN_4O_{12}$ requires 819.2639.

(+)-(1S)-1-[(1R)-1-Chloroethyl]-7-cyano-3- $({5}$ -[2-(dimethylamino)ethoxy|indol-2-yl}carbonyl)-1,2-dihydro-3H-benz[e|indol-5-yl 2,3,4,6-Tetra-O-acetyl-β-D-galactopyranoside [(+)-(1S,10R)-27b]: A mixture of the phenol (+)-(1S,10R)-24b (218 mg, 585 μmol, 1.00 equiv.) in dry CH₂Cl₂ (29 mL) and freshly activated molecular sieves (4 Å, 1.50 g) was stirred at room temperature for 30 min. After addition of the trichloroacetimidate 25 (317 mg, 643 µmol, 1.1 equiv.) and cooling to -10 °C, the promoter BF₃·OEt₂ (37 µL, 292 µmol, 0.5 equiv.) in dry CH₂Cl₂ (2.9 mL) was added dropwise, and the mixture was stirred for 3.5 h. Additional BF₃·OEt₂ (222 µL, 1.75 mmol, 3.0 equiv.) in CH₂Cl₂ (2.8 mL) was then added, and the mixture was allowed to warm to room temperature and stirred for 6 h. The reaction mixture was transferred by cannula into a new flask for separation of the molecular sieves, which were washed with CH₂Cl₂ (2 × 20 mL). The obtained organic solution was concentrated in vacuo, and the residue was dissolved in DMF (29 mL) and cooled to 0 °C. EDC·HCl (336 mg, 1.75 mmol, 3.0 equiv.) and DMAI·HCl (26, 250 mg, 878 µmol, 1.5 equiv.) were added, and the reaction mixture was stirred at room temperature for 18 h. After dilution with EtOAc (50 mL), H₂O (50 mL) and saturated aqueous NaHCO₃ (50 mL) were added, and the aqueous phase was extracted with EtOAc (4× 100 mL). The combined organic layers were washed with brine (4× 200 mL) and dried (MgSO₄) and concentrated in vacuo. Purification of the residue by column chromatography (SiO₂; CH₂Cl₂/MeOH, 10:1) afforded (+)-(1S,10R)-27b as a white solid (254 mg, 305 µmol, 52%). $[a]_D^{20} =$ +6.7 (c = 0.3 in CHCl₃). ¹H NMR (600 MHz, CDCl₃): $\delta = 1.63$ $(d, J = 6.7 \text{ Hz}, 3 \text{ H}, 11\text{-H}_3), 2.00 \text{ [s, 3 H, C(O)CH}_3], 2.02 \text{ [s, 3 H, C(O)CH}_3]$ C(O)CH₃], 2.06 [s, 3 H, C(O)CH₃], 2.19 [s, 3 H, C(O)CH₃], 2.39 (s,

6 H, NMe₂), 2.81 (t, J = 5.6 Hz, 2 H, 2"-H₂), 4.02–4.07 (m, 2 H, 1-H, 5'''-H), 4.11–4.23 (m, 4 H, 1''-H $_2$, 6'''-H $_2$), 4.52 (m $_c$, 1 H, 10-H), 4.63 (m_c, 1 H, 2-H_a), 4.86 (dd, J = 10.6, 2.1 Hz, 1 H, 2-H_b), 5.10 (dd, J = 10.5, 3.5 Hz, 1 H, 3'''-H), 5.30 (d, J = 8.0 Hz, 1 H, 1'''-H), 5.44 (m_c, 1 H, 4'''-H), 5.66 (dd, J = 10.5, 8.0 Hz, 1 H, 2'''-H), 7.01-7.06 (m, 2 H, 3'-H, 6'-H), 7.13 (d, J = 2.3 Hz, 1 H, 4'-H), 7.36 (d, J = 8.9 Hz, 1 H, 7'-H), 7.62 (dd, J = 8.7, 1.5 Hz, 1 H, 8-H), 7.75 (d, J = 8.7 Hz, 1 H, 9-H), 8.43 (br. s, 1 H, 4-H), 8.51 (d, J = 1.5 Hz, 1 H, 6-H), 9.62 (br. s, 1 H, NH) ppm. ¹³C NMR (150 MHz, CDCl₃): $\delta = 20.6$ (3 signals), 20.7 [4× C(O)*C*H₃], 23.69 (C-11), 45.74 (NMe₂), 47.15 (C-1), 53.06 (C-2), 58.25 (C-2''), 59.32 (C-10), 61.09 (C-6'''), 66.26 (C-1''), 66.74 (C-4'''), 68.46 (C-2'''), 70.58 (C-3'''), 71.19 (C-5'''), 99.44 (C-1'''), 103.3, 103.5 (C-4, C-4'), 106.3 (C-3'), 107.4 (C-7), 112.8 (C-7'), 117.7 (C-6'), 119.2, 119.4, 122.6 (C-5a, C-9b, CN), 123.6 (C-9), 128.1, 128.2 (C-8, C-3a'), 129.8, 130.1, 131.1, 131.6 (C-6, C-9a, C-2', C-7a'), 145.3 (C-3a), 153.7, 153.9 (C-5, C-5'), 160.7 (C=O), 169.4, 170.0, 170.2, 170.5 [4× C(O)CH₃] ppm. IR (KBr): \tilde{v} = 3406 (NH), 2940, 2226 (CN), 1752 (C=O), 1624, 1521, 1457 cm⁻¹. UV (CH₃CN): λ_{max} (lg ε) = 218.0 (4.656), 266.5 (4.415), 309.0 (4.487), 359.0 (4.540) nm. ESI-HRMS: $m/z = 833.2795 [M + H]^+$; $C_{42}H_{45}ClN_4O_{12}$ requires 833.2796.

(+)-(1S)-8-(Aminomethyl)-1-(chloromethyl)-3-({5-[2-(dimethylamino)ethoxy|indol-2-yl}carbonyl)-1,2-dihydro-3H-benz[e|indol-5-yl 2,3,4,6-Tetra-O-acetyl-β-D-galactopyranoside [(+)-(1S)-28a]: A solution of (+)-(1S)-27a (50 mg, 61 μ mol, 1.0 equiv.) in anhydrous EtOH (3 mL) was flushed with argon, and a solution of HCl in EtOH (1.25 M, 98 μL, 122 μmol, 2.0 equiv.), H₂O (0.15 mL) and PtO₂·H₂O (3.0 mg, 12.2 μmol, 0.2 equiv.) were added. The mixture was stirred at room temperature under H₂ for 48 h and then filtered through Celite, with washing with MeOH (200 mL), and concentrated in vacuo. Purification of the residue by column chromatography (SiO₂; CH₂Cl₂/MeOH, 5:1) afforded (+)-(1S)-28a as a white solid (33 mg, 40 μ mol, 66%). [a]_D²⁰ = +2.0 (c = 0.3 in DMSO). ¹H NMR (600 MHz, $[D_6]DMSO$): $\delta = 1.98$ [s, 3 H, C(O)CH₃], 2.01 [s, 3 H, C(O)CH₃], 2.04 [s, 3 H, C(O)CH₃], 2.18 [s, 3 H, C(O)CH₃], 2.24 (s, 6 H, NMe₂), 2.65 (t, J = 5.8 Hz, 2 H, 2"-H₂), 3.89–3.97 (m, 3 H, CH_2NH_2 , 10-H_a), 4.07 (t, J = 5.9 Hz, 2 H, 1"-H₂), 4.14 (m, 3 H, $6^{\prime\prime\prime}$ -H₂, 10-H_b), 4.26 (m, 1 H, 1-H), 4.49 (m, 1 H, $5^{\prime\prime\prime}$ -H), 4.57–4.62 (m, 1 H, 2-H_a), 4.83 (m, 1 H, 2-H_b), 5.38–5.46 (m, 3 H, 2'''-H, 3'''-H, 4'''-H), 5.56 (m, 1 H, 1'''-H), 6.92 (dd, J = 8.9, 2.3 Hz, 1 H, 6'-H), 7.10 (s, 1 H, 3'-H), 7.17 (d, J = 2.1 Hz, 1 H, 4'-H), 7.41 (d, J = 8.9 Hz, 1 H, 7'-H), 7.44 (d, J = 8.7 Hz, 1 H, 7-H), 7.86 (s, 1 H, 9-H), 7.90 (d, J = 8.7 Hz, 1 H, 6-H), 8.18 (s, 1 H, 4-H), 11.55 (s, 1 H, NH) ppm. 13 C NMR (125 MHz): $\delta = 20.29$, $20.34, 20.50 [4 \times C(O)CH_3], 40.09 (C-1), 41.14 (C-10), 45.51$ (CH₂NH₂), 47.42 (NMe₂), 55.01 (C-2), 57.76 (C-2''), 60.94 (C-1''), 5'''), 98.90 (C-1'''), 101.6 (C-4), 103.1 (C-4'), 105.2 (C-3'), 113.1 (C-7'), 115.7 (C-6'), 118.8, 121.2, 127.3, 129.5, 130.7, 131.5 (C-5a, C-9a, C-9b, C-8, C-2', C-3a', C-7a'), 121.6 (C-6), 124.5 (C-7), 141.8 (C-3a), 152.6, 152.8 (C-5, C-5'), 160.1 (C=O), 169.2, 169.3, 169.7, 169.8 [4 × C(O)CH₃] ppm. IR (KBr): \tilde{v} = 3421 (NH), 1749 (C=O), 1628, 1519, 1404 cm⁻¹. UV (CH₃CN): λ_{max} (lg ε) = 206.0 (4.461), 300.0 (4.312), 336.0 (4.249) nm. ESI-HRMS: m/z = 823.2941 [M + H_1^+ ; $C_{41}H_{47}C1N_4O_{12}$ requires 823.2952.

(+)-(1*S*)-7-(Aminomethyl)-1-[(1*R*)-1-chloroethyl]-3-({5-[2-(dimethylamino)ethoxy|indol-2-yl}carbonyl)-1,2-dihydro-3*H*-benz[e|indol-5-yl 2,3,4,6-Tetra-O-acetyl- β -D-galactopyranoside [(+)-(1*S*,10*R*)-28b]: A solution of (+)-(1*S*,10*R*)-27b (50 mg, 60 μmol, 1.0 equiv.) in anhydrous EtOH (3 mL) was flushed with argon, and a solution of HCl in EtOH (1.25 M, 96 μL, 120 μmol, 2.0 equiv.), H₂O (0.15 mL) and PtO₂·H₂O (3.0 mg, 12.0 μmol, 0.2 equiv.) were added. The mixture



was stirred at room temperature under H₂ for 48 h and was then filtered through Celite, with washing with MeOH (100 mL), and concentrated in vacuo. Purification of the residue by column chromatography (SiO₂; CH₂Cl₂/MeOH, 5:1) afforded (+)-(1S,10R)-**28b** as a white solid (41 mg, 49 μ mol, 82%). [a]²⁰ = +4.4 (c = 0.25 in DMSO). ¹H NMR (600 MHz, [D₆]DMSO): δ = 1.64 (d, J = 6.3 Hz, 3 H, 11-H₃), 1.98, 1.99, 2.06, 2.18 [$4 \times s$, 12 H, $4 \times C(O)$ CH₃], 2.25 (m_c, 6 H, NMe₂), 2.67 (m_c, 2 H, 2"-H₂), 3.93-4.19 (m, 6 H, 1"-H₂, 6"-H₂, CH₂NH₂), 4.27 (m_c, 1 H, 1-H), 4.45–4.53 (m, 1 H, 5'''-H), 4.58–4.65 (m, 1 H, 2-H_a), 4.73–4.84 (m, 2 H, 2-H_b, 10-H), 5.35-5.48 (m, 3 H, 2'''-H, 3'''-H, 4'''-H), 5.59 (d, J = 7.5 Hz, 1H, 1'''-H), 6.92 (m_c, 1 H, 6'-H), 7.14–7.20 (m, 2 H, 3'-H, 4'-H), 7.40–7.44 (m, 1 H, 7'-H), 7.72–7.79 (m, 1 H, 6-H), 7.93 (br. s, 1 H, 9-H*), 8.03 (d, J = 8.6 Hz, 1 H, 8-H*), 8.24 (br. s, 1 H, 4-H), 11.57 $(m_c, 1 \text{ H}, \text{ NH}) \text{ ppm.}^{-13}\text{C NMR} (125 \text{ MHz}, [D_6]\text{DMSO}): \delta = 20.30$ (3 signals), 20.60 [$4 \times C(O)CH_3$], 23.29 (C-11), 44.29 (CH_2NH_2), 45.45 (NMe₂), 45.92 (C-1), 52.07 (C-2), 57.74 (C-2"), 61.03, 61.25 (C-10, C-6'''), 66.24 (C-1''), 67.14, 68.66, 69.89 (C-2''', C-3''', C-3''') 4'''), 70.43 (C-5'''), 99.16 (C-1'''), 102.8 (C-4), 103.3 (C-4'), 105.6 (C-3'), 113.2 (C-7'), 115.9 (C-6'), 120.2, 121.4, 122.4, 123.5 (C-7, C-8, C-9, C-9b*), 127.5, 127.9, 128.8, 130.7, 131.8, 134.5 (C-6, C-5a*, C-9a, C-2', C-3a', C-7a'), 141.9 (C-3a), 152.6, 153.0 (C-5, C-5'), 160.2 (C=O), 169.4, 169.6, 169.9, 170.0 [4× C(O)CH₃] ppm. IR (KBr): $\tilde{v} = 3417$ (NH), 1750 (C=O), 1624, 1520, 1471, 1409 cm⁻¹. UV (CH₃CN): λ_{max} (lg ε) = 202.5 (4.555), 249.0 (4.248), 298.0 (4.343), 337.5 (4.331) nm. ESI-HRMS: m/z = 837.3108 [M + H]+; C₄₂H₄₉ClN₄O₁₂ requires 837.3109.

(1S)-8-(Aminomethyl)-1-(chloromethyl)-3-({5-[2-(dimethylamino)ethoxy|indol-2-yl}carbonyl)-1,2-dihydro-3*H*-benz[*e*|indol-5-yl β-D-Galactopyranoside [(1S)-3a]: A solution of NaOMe in MeOH (5.4 M, 8.5 μL, 46 μmol, 2.0 equiv.) was added at 0 °C to a solution of (+)-(1S)-28a (19 mg, 23 μmol, 1.0 equiv.) in MeOH (3 mL). The mixture was allowed to warm to room temperature and stirred for 30 min. MeOH (1 mL), H₂O (1 mL) and Amberlite IR-120 were then added (pH = 7). The mixture was filtered, the filter cake was washed with MeOH (25 mL), and the filtrate was concentrated in vacuo. The residue was dissolved in H₂O + 0.1% CF₃CO₂H/ CH₃CN (4:1) (4 mL) and purified by prep. HPLC to afford 3a as a white solid (12.7 mg, 19.4 μ mol, 84%). ¹H NMR (600 MHz, [D₆]-DMSO): $\delta = 2.90$ (s, 6 H, NMe₂), 3.44–4.21 (m, 7 H, 1"-H, 2"-H, 3'''-H, 4'''-H, 5'''-H, 6'''-H), 4.20–4.31 (m, 3 H, $\mathrm{C}H_2\mathrm{NH}_2$, 1-H), 4.31-4.40 (m, 2 H, 2"-H₂), 4.57-4.66 (m, 2 H, 10-H₂), 4.82-4.93 (m, 3 H, 2-H₂, 1'''-H), 7.01 (dd, J = 8.9, 2.4 Hz, 1 H, 6'-H),7.14 (d, J = 1.6 Hz, 1 H, 3'-H), 7.26 (d, J = 2.3 Hz, 1 H, 4'-H),7.45 (d, J = 8.9 Hz, 1 H, 7'-H), 7.49 (dd, J = 8.8, 1.3 Hz, 1 H, 7-H), 8.01 (s, 1 H, 9-H), 8.37 (d, J = 8.7 Hz, 1 H, 6-H), 8.42 (s, 1 H, 4-H), 11.74 (s, 1 H, NH) ppm. ¹³C NMR (125 MHz, [D₆]DMSO): $\delta = 41.29 \text{ (C-1)}, 42.30 \text{ (CH}_2\text{NH}_2), 42.80 \text{ (NMe}_2), 47.27 \text{ (C-10)},$ 54.89 (C-2), 55.55 (C-1''), 59.45 (C-6'''), 62.72 (C-2''), 67.40, 70.38, 73.02, 75.05 (C-2", C-3", C-4", C-5"), 102.1 (C-1"), 103.9 (C-4'), 105.2 (C-3'), 113.2 (C-7'), 115.6 (C-6'), 117.8 (C-4*), 122.3 (C-9b), 122.5 (C-9), 123.7 (C-6), 124.1 (C-7), 127.2, 129.1, 131.0, 131.9, 133.1 (C-5a, C-9a, C-2', C-3a', C-7a', C-8), 142.5 (C-3a), 151.9, 153.6 (C-5, C-5'), 160.1 (C=O) ppm. ESI-HRMS: m/z = $655.2526 [M + H]^+$; $C_{33}H_{39}ClN_4O_8$ requires 655.2529.

(-)-(1*S*)-7-(Aminomethyl)-1-[(1*R*)-1-chloroethyl]-3-({5-[2-(dimethyl-amino)ethoxy|indol-2-yl}carbonyl)-1,2-dihydro-3*H*-benz[e|indol-5-yl}β-D-Galactopyranoside Bis(trifluoroacetate) [(-)-(1*S*,10*R*)-3b]: A solution of NaOMe in MeOH (5.4 m, 44 μL, 239 μmol, 2.0 equiv.) was added at 0 °C to a solution of (+)-(1*S*,10*R*)-28b (100 mg, 119 μmol, 1.0 equiv.) in MeOH (12 mL). The reaction mixture was allowed to warm to room temperature and stirred for 30 min. MeOH (4 mL), H₂O (4 mL) and Amberlite IR-120 were added (pH

= 7). The reaction mixture was filtered, the filter cake was washed with MeOH (120 mL), and the filtrate was concentrated in vacuo. The residue was dissolved in $H_2O + 0.1\% CF_3CO_2H/CH_3CN$ (4:1) (12 mL) and purified by prep. HPLC to afford (-)-(1S,10R)-3b as a white solid (75 mg, 84 μ mol, 70%). [a]_D²⁰ = -9.0 (c = 0.3 in DMSO). ¹H NMR (600 MHz, [D₆]DMSO, 100 °C): δ = 1.65 (d, J = 6.5 Hz, 3 H, 11-H₃), 2.90 (s, 6 H, NMe₂), 3.48–3.55, 3.56–3.64 $(2 \times m, 5 H, 2''-H_2, 3'''-H, 5'''-H, 6'''-Ha), 3.67-3.73 (m, 1 H,$ 6'''-H_b), 3.82-3.89 (m, 2 H, 2'''-H, 4'''-H), 4.18-4.29 (m, 3 H, 1-H, $CH_2NH_3^+$), 4.36 (m_c, 2 H, 1''-H₂), 4.62–4.67 (m, 1 H, 2-H_a), $4.74 \text{ (m}_c, 1 \text{ H}, 2-\text{H}_b), 4.82 \text{ (m}_c, 1 \text{ H}, 10-\text{H}), 4.97 \text{ (d}, J = 7.6 \text{ Hz}, 1)$ H, 1'''-H), 7.02 (m_c, 1 H, 6'-H), 7.15 (d, J = 1.8 Hz, 1 H, 3'-H), 7.28 (br. s, 1 H, 4'-H), 7.47 (d, J = 8.9 Hz, 1 H, 7'-H), 7.64 (d, J= 8.8 Hz, 1 H, 8-H), 8.03 (d, J = 8.6 Hz, 1 H, 9-H), 8.24 (br. s, 1)H, 4-H), 8.41 (br. s, 1 H, 6-H), 11.52 (s, 1 H, NH) ppm. ¹³C NMR (125 MHz, [D₆]DMSO): δ = 23.28 (C-11), 42.65 (CH₂NH₃⁺), 42.84 (NMe₂), 45.89 (C-1), 52.03 (C-2), 55.60 (C-2"), 59.60 (C-6""), 61.27 (C-10), 62.81 (C-1"), 67.48, 70.56 (C-2", C-4"), 73.01, 75.20 (C-3''', C-5'''), 101.9 (C-4), 102.2 (C-1'''), 104.1 (C-4'), 105.5 (C-3'), 113.3 (C-7'), 115.7 (C-6'), 118.8, 122.5 (C-7, C-9b), 123.6 (C-9), 123.8 (C-6), 127.4, 127.7, 129.1, 129.2, 131.1, 132.1 (C-5a, C-8, C-9a, C-2', C-3a', C-7a'), 142.6 (C-3a), 152.0, 153.7 (C-5, C-5'), 158.1 [q, ${}^{2}J_{C,F} = 32 \text{ Hz}$, CF₃C(O)], 160.1 (C=O) ppm. IR (KBr): $\tilde{v} = 3406$ (NH), 1679, 1624, 1521, 1472, 1412 cm⁻¹. UV (CH₃CN): λ_{max} (lg ε) = 207.5 (4.416), 250.5 (4.158), 299.0 (4.247), 337.5 nm (4.230). ESI-HRMS: $m/z = 669.2686 \text{ [M} - 2 \times \text{CF}_3\text{CO}_2 - \text{CF}_3\text{CO}_2 + \text{CF}_3\text{CO}_2 - \text{CF}_3\text{CO}_2 - \text{CF}_3\text{CO}_2 + \text{CF}_3\text{CO}_2 - \text{CF}_3\text{CO}_2 -$ H]⁺; C₃₈H₄₃ClF₆N₄O₁₂ requires 669.2685.

Synthesis of the Fluorescein-Labelled Compound (1S)-30a: A solution of 3a (11.9 mg, 18.2 μ mol, 1.3 equiv.) and iPr_2NEt (11 μ L, $14.5 \,\mu g$, $66 \,\mu mol$, $4.5 \,equiv$.) in anhydrous DMF ($150 \,\mu L$) was added to a solution of 29 (8.90 mg, 15.2 µmol, 1.0 equiv.) in anhydrous DMF (150 µL), and the mixture was stirred at room temperature for 18 h; then H₂O + 0.1% CF₃CO₂H (3.0 mL) and CH₃CN (2.5 mL) were added, and the compound was purified by prep. HPLC to afford (1S)-30a as a yellow solid (7.8 mg, 6.9 µmol, 46%). ¹H NMR (600 MHz, [D₆]DMSO): δ = 1.08–1.70 (m, 6 H, 3^S-H₂, 4^S-H₂, 5^S-H₂), 2.20 (m, 2 H, 2^S-H), 2.87 (s, 6 H, NMe₂), 3.18–4.26 (m, 13 H, 2'''-H, 3'''-H, 4'''-H, 5'''-H, 6'''-H₂, 10-H₂, 1-H, 6^{S} -H₂, 1''-H₂), 4.37 (m, 2 H, 2''-H), 4.47 (m, 2 H, Ar-CH₂), 4.59 (m, 1 H, 2-H_a), 4.85 (m, 1 H, 2-H_b), 4.90 (m, 1 H, 1'''-H), 6.53-6.60 (m, 4 H, Ar-H), 6.69 (d, J = 2.1 Hz, 1-H, Ar-H), 7.01(dd, J = 8.9, 2.4 Hz, 1 H, 6'-H), 7.18 (m, 1 H, 3'-H), 7.27 (d, J =2.1 Hz, 1 H, 4'-H), 7.30–7.38 (m, 2 H, 7'-H, Ar-H), 7.45 (d, J =8.9 Hz, 1 H, 7-H), 7.72 (m, 1 H, Ar-H), 8.24 (m, 1 H, Ar-H), 8.30 (d, J = 8.8 Hz, 1 H, 6-H), 8.41 (t, J = 6.0 Hz, 1 H, Ar-H), 8.45 (d, J = 9.6 Hz, 1 H, Ar-H), 8.79 (t, J = 5.4 Hz, 1 H, Ar-H), 9.75, 10.05 (s, 2 H, OH, CO₂ H), 11.70 (s, 1 H, indole-NH) ppm. ESI-HRMS: $m/z = 1126.3832 \text{ [M + H]}^+; C_{60}H_{60}ClN_5O_{15} \text{ requires } 1126.3847.$

Chromatographic Resolution of (1*S*)-30a: Compound (1*S*)-30a was separated consecutively (injection volume: 2.0 mL) by semipreparative HPLC [Kromasil 100 C18; 250 × 20 mm; particle size: 7 µm; gradient: H₂O (0.1% CF₃CO₂H)/CH₃CN, 83:17 for 60 min, then H₂O (0.1% CF₃CO₂H)/CH₃CN, 50:50 for 10 min, then H₂O (0.1% CF₃CO₂H)/CH₃CN, 83:17 for 10 min; flow: 12 mL min⁻¹; UV detector: λ = 250 nm] to provide (1*S*)-30a (t_R = 33.1 min).

Synthesis of the Fluorescein-Labelled Compound (1*S*,10*R*)-30b: A solution of (–)-(1*S*,10*R*)-3b (11.95 mg, 13.3 µmol, 1.0 equiv.) and iPr_2NEt (11 µL, 14.5 µg, 66 µmol, 5.0 equiv.) in anhydrous DMF (150 µL) was added to a solution of **29** (8.28 mg, 14.1 µmol, 1.0 equiv.) in anhydrous DMF (150 µL), and the mixture was stirred at room temperature for 19 h; then $H_2O + 0.1\%$ CF₃CO₂H (3.0 mL) and CH₃CN (2.7 mL) were added, and the compound was

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purified by prep. HPLC to afford (1S,10R)-30b as a yellow solid (14.7 mg, 11.7 μmol, 88%). ¹H NMR (600 MHz, [D₆]DMSO, 80 °C): $\delta = 1.37-1.45$ (m, 2 H, 4^S-H₂), 1.57-1.69 (m, 7 H, 11-H₃, $3^{S}-H_{2}$, $5^{S}-H_{2}$), 2.22 (m_c, 2 H, $2^{S}-H_{2}$), 2.92 (s, 6 H, NMe₂), 3.34 (m_c, 2 H, 6^S-H₂), 3.47-3.63 (m, 5 H, 2"-H₂, 3"-H, 5"-H, 6"-H_a), 3.67-3.72 (m, 1 H, 6"'-H_b), 3.81-3.87 (m, 2 H, 2"-H, 4"-H), $4.22 \text{ (m}_c, 1 \text{ H}, 1\text{-H}), 4.36 \text{ (m}_c, 2 \text{ H}, 1''\text{-H}_2), 4.45 \text{ (m}_c, 2 \text{ H}, ArCH_2),}$ 4.63 (m_c, 1 H, 2-H_a), 4.71 (m_c, 1 H, 2-H_b), 4.81 (m_c, 1 H, 10-H), 4.93 (d, J = 7.5 Hz, 1 H, 1'''-H), 6.56 (m_c, 4 H, 4× Ar-H), 6.70 (br. s, 2 H, $2 \times$ Ar-H), 7.02 (m_c, 1 H, 6'-H), 7.13 (d, J = 1.5 Hz, 1 H, 3'-H), 7.25–7.35, 7.44–7.51 ($2 \times m$, 4 H, 8-H, 4'-H, 7'-H, Ar-H), 7.91 (d, J = 8.7 Hz, 1 H, 9-H), 8.07–8.27 (m, 4 H, 4-H, $2 \times$ Ar-H, NH), 8.44 (br. s, 1 H, 6-H), 8.58 (br. s, 1 H, NH), 9.89, 9.59 (2× br. s, 2 H, OH, CO₂ H), 11.51 (br. s, 1 H, indole-NH) ppm. ESI-HRMS: $m/z = 1140.4004 [M - 2 \times CF_3CO_2]^+$; $C_{63}H_{63}ClF_{3}$ -N₅O₁₇ requires 1140.4004.

Chromatographic Purification of (1*S*,10*R*)-30b: Semipreparative HPLC [Kromasil 100 C18; 250 × 20 mm; particle size: 7 μ m; gradient: H₂O (0.1% CF₃CO₂H)/CH₃CN, 83:17 for 60 min, then H₂O (0.1% CF₃CO₂H)/CH₃CN, 50:50 for 10 min, then H₂O (0.1% CF₃CO₂H)/CH₃CN, 83:17 for 10 min; flow: 12 mL min⁻¹; UV detector: $\lambda = 250$ nm; injection volume: 2.0 mL]. (1*S*)-30a ($t_R = 36.6$ min).

Synthesis of the Dapoxyl-Labelled Compound (-)-(1S,10R)-32: A solution of (-)-(1S,10R)-3b (25.0 mg, 27.9 µmol, 1.0 equiv.) and iPr_2NEt (23 µL, 30.3 µg, 139 µmol, 5.8 equiv.) in anhydrous DMF $(150 \,\mu\text{L})$ was added to a solution of 31 (12.3 mg, 24.0 μ mol, 1.0 equiv.) in anhydrous DMF (200 µL), and the mixture was stirred at room temperature for 9 h. H₂O (+ 0.1% CF₃CO₂H) (3.0 mL) and CH₃CN (2.5 mL) were added, and the compound was purified by prep. HPLC to afford (-)-(1S,10R)-32 as a yellow solid (27.8 mg, 21.5 μmol, 89%). ¹H NMR (600 MHz, [D₆]DMSO, 80 °C): $\delta = 1.63$ (d, J = 6.6 Hz, 3 H, 11-H₃), 2.40 (t, J = 7.4 Hz, 2 H, 2^S-H₂), 2.93 (s, 6 H, 2"-NMe₂), 2.98 (s, 6 H, 12#-NMe₂), 3.14 $(t, J = 7.4 \text{ Hz}, 2 \text{ H}, 3^{\text{S}} - \text{H}_2), 3.48 \text{ (dd}, J = 9.6, 3.2 \text{ Hz}, 1 \text{ H}, 3''' - \text{H}),$ 3.52-3.57 (m, 3 H, 5'''-H, 2''-H₂), 3.60 (dd, J = 10.6, 5.5 Hz, 1 H, $6'''-H_a$, 3.69 (dd, J = 10.6, 7.2 Hz, 1 H, $6'''-H_b$), 3.81–3.86 (m, 2 H, $2^{\prime\prime\prime}$ -H, $4^{\prime\prime\prime}$ -H), 4.20 (dt, J = 9.4, 2.6 Hz, 1 H, 1-H), 4.36 (t, J= 5.0 Hz, 2 H, 1''-H₂), 4.42 (m_c, 2 H, 12-H₂), 4.62 (dd, J = 11.0, 2.6 Hz, 1 H, 2-H_a), 4.70 (dd, J = 11.0, 9.4 Hz, 1 H, 2-H_b), 4.79(dq, J = 6.6, 3.0 Hz, 1 H, 10-H), 4.92 (d, J = 7.8 Hz, 1 H, 1'''-H),6.83 (d, J = 9.0 Hz, 2 H, 2× Ar-H), 7.01 (dd, J = 9.0, 2.2 Hz, 1 H, 6'-H), 7.12 (d, J = 2.0 Hz, 1 H, 3'-H), 7.28 (d, J = 2.2 Hz, 1 H, 4'-H), 7.45-7.48 (m, 2 H, 7'-H, 8-H), 7.54-7.56 (m, 2 H, 3^S-NH, 8[#]-H), 7.65 (d, J = 9.0 Hz, 2 H, 2× Ar-H), 7.89 (d, J = 8.5 Hz, 1 H, 9-H), 7.96 (d, J = 8.4 Hz, 2 H, 2× Ar-H), 8.18 (s, 1 H, 12-NH), 8.20-8.23 (m, 4 H, 4-H, 6-H, $2 \times Ar-H$), 9.64 (br. s, 1 H, NH^+), 11.5 (s, 1 H, 1'-NH) ppm. ¹³C NMR (125 MHz, [D₆]DMSO): δ = 23.4 (C-11), 35.5 (C-2^S), 38.7 (C-3^S), 40.1 (12[#]-NMe₂), 42.5 (C-12), 42.9 (2"-NMe₂), 46.0 (C-1), 52.0 (C-2), 55.7 (C-2"), 59.5 (C-6""), 61.4 (C-10), 62.7 (C-1"), 67.5 (C-4""), 70.5 (C-2""), 73.1 (C-3""), 75.2 (C-5'''), 101.8 (C-9b), 102.1 (C-1'''), 104.0 (C-4'), 105.4 (C-3'), 112.1 (2 × Ar-C), 113.3 (C-7'), 114.6 (CF₃CO₂), 115.6 (C-6'), 118.9 (C-5a), 121.4 (C-8#), 121.7 (C-4, C-6), 122.7 (C-7), 123.3 (C-9), 125.5 (2 × Ar-C), 126.1 (2 × Ar-C), 127.4 (C-8), 127.4 (2 × Ar-C), 128.5 (C-4#), 130.2 (C-9#), 131.2 (C-9a), 132.0 (C-3a'), 134.7 (C-7a'), 141.1 (C-2', C-1#), 141.6 (C-3a), 150.1 (C-7#), 152.0 (C-12#), 152.7 (C-5'), 153.4 (C-5), 157.5 (C-5#, 1'-C=O), 160.0 (CF_3CO_2) , 169.6 (2^S-C=O) ppm. IR (KBr): $\tilde{v} = 3406$, 2924, 2819, 1675, 1616, 1510, 1407, 1161, 800 cm⁻¹. UV (MeOH): λ_{max} (lg ε) = 202.5 (4.821), 252.0 (4.514), 289.0 (4.546), 299.5 (4.582), 349.0 nm (4.627). ESI-HRMS: m/z = 1066.3778 [M - $2 \times \text{CF}_3\text{CO}_2$]⁺; C₅₆H₆₁ClF₃N₇O₁₄S requires 1066.3782.

Chromatographic Purification of (–)-(1*S***,10***R***)-32: Semipreparative HPLC [Kromasil 100 C18; 250 × 20 mm; particle size: 7 μm; gradient: H₂O (0.1% CF₃CO₂H)/CH₃CN, 83:17 for 60 min, then H₂O (0.1% CF₃CO₂H)/CH₃CN, 50:50 for 10 min, then H₂O (0.1% CF₃CO₂H)/CH₃CN, 83:17 for 10 min; flow: 12 mL min⁻¹; UV detector: \lambda = 250 nm; injection volume: 2.0 mL]. (1***S***,10***R***)-32 (t_R = 38.5 min). [a]^D₀ = −14.0 (c = 0.15, MeOH).**

Supporting Information (see footnote on the first page of this article): Experimental Section for the synthesis of molecules 7a–17c.

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- [1] a) L. F. Tietze, B. Krewer, F. Major, I. Schuberth, J. Am. Chem. Soc. 2009, 131, 13031-13036; b) L. F. Tietze, B. Krewer, J. M. von Hof, H. Frauendorf, I. Schuberth, Toxins 2009, 1, 134-150; c) L. F. Tietze, B. Krewer, Anti-Cancer Agents Med. Chem. **2009**, 9, 304–325; d) L. F. Tietze, B. Krewer, H. Frauendorf, F. Major, I. Schuberth, Angew. Chem. 2006, 118, 6720-6724; Angew. Chem. Int. Ed. 2006, 45, 6570-6574; e) L. F. Tietze, T. Feuerstein, A. Fecher, F. Haunert, O. Panknin, U. Borchers, I. Schuberth, F. Alves, Angew. Chem. 2002, 114, 785-787; f) L. F. Tietze, T. Feuerstein, A. Fecher, F. Haunert, O. Panknin, U. Borchers, I. Schuberth, F. Alves, Angew. Chem. Int. Ed. 2002, 41, 759–761; g) L. F. Tietze, M. Lieb, T. Herzig, F. Haunert, I. Schuberth, Bioorg. Med. Chem. 2001, 9, 1929-1939; h) L. F. Tietze, R. Hannemann, W. Buhr, M. Lögers, P. Menningen, M. Lieb, D. Starck, T. Grote, A. Döring, I. Schuberth, Angew. Chem. 1996, 108, 2840–2842; i) L. F. Tietze, R. Hannemann, W. Buhr, M. Lögers, P. Menningen, M. Lieb, D. Starck, T. Grote, A. Döring, I. Schuberth, Angew. Chem. Int. Ed. Engl. **1996**, *35*, 2674–2677.
- [2] K. D. Bagshawe, Br. J. Cancer 1987, 56, 531–532.
- [3] a) Reviews: L. F. Tietze, B. Krewer, Chem. Biol. Drug Des. 2009, 74, 205–211; b) K. D. Bagshawe, Curr. Drug Targets 2009, 10, 152–157; c) K. D. Bagshawe, Expert Rev. Anticancer Ther. 2006, 6, 1421–1431; d) W. A. Denny, Cancer Invest. 2004, 22, 604–619; e) L. F. Tietze, T. Feuerstein, Curr. Pharm. Des. 2003, 9, 2155–2175; f) L. F. Tietze, T. Feuerstein, Aust. J. Chem. 2003, 56, 841–854; g) M. Jung, Mini-Rev. Med. Chem. 2001, 1, 399–407; h) G. Xu, H. L. McLeod, Clin. Cancer Res. 2001, 7, 3314–3324; i) K. N. Syrigos, A. A. Epenetos, Anticancer Res. 1999, 19, 605–613; j) G. M. Dubowchik, M. A. Walker, Pharmacol. Ther. 1999, 83, 67–123; k) I. Niculescu-Duvaz, C. J. Springer, Adv. Drug Delivery Rev. 1997, 26, 151–172; l) L. N. Jungheim, T. A. Shepherd, Chem. Rev. 1994, 94, 1553–1566.
- [4] a) L. J. Hanka, A. Dietz, S. A. Gerpheide, S. L. Kuentzel, D. G. Martin, J. Antibiot. 1978, 31, 1211–1217; b) D. G. Martin, C. Biles, S. A. Gerpheide, L. J. Hanka, W. C. Krueger, J. P. McGovren, S. A. Mizsak, G. L. Neil, J. C. Stewart, J. Visser, J. Antibiot. 1981, 34, 1119–1125.
- [5] a) L. F. Tietze, F. Major, I. Schuberth, Angew. Chem. 2006, 118, 6724–6727; Angew. Chem. Int. Ed. 2006, 45, 6574–6577;
 b) L. F. Tietze, F. Major, I. Schuberth, D. A. Spiegl, B. Krewer, K. Maksimenka, G. Brinkmann, J. Magull, Chem. Eur. J. 2007, 13, 4396–4409.
- [6] a) L. D. Lavis, T.-Y. Chao, R. T. Raines, ACS Chem. Biol. 2006,
 1, 252–260; b) A. Miyawaki, A. Sawano, T. Kogure, Nat. Cell Biol. 2003, 5, 1–7; c) J. Lippincott-Schwartz, G. Patterson, Science 2003, 300, 87–91; d) D. B. Zorov, E. Kobrinsky, M. Juhaszova, S. J. Sollott, Circ. Res. 2004, 95, 239–252.



- [7] C. Dullin, M. Zientkowska, J. Napp, J. Missbach-Guentner, H.-W. Krell, F. Müller, E. Grabbe, L. F. Tietze, F. Alves, *Imaging* 2009, 8, 1–14.
- [8] a) C. O. Kappe, D. Dallinger, Mol. Diversity 2009, 13, 71–193;
 b) J. A. Corner, S. W. Luning, R. Price, J. Chem. Soc. Perkin Trans. 1 1990, 1127–1132;
 c) G. P. Ellis, T. M. Romney-Alexander, Chem. Rev. 1987, 87, 779–794;
 d) J. Linley, Tetrahedron 1984, 40, 1433–1456.
- [9] L. F. Tietze, H. J. Schuster, J. Marian von Hof, S. M. Hampel, J. F. Colunga, M. John, *Chem. Eur. J.*, in press.
- [10] Compound 22 had already been prepared by a slightly different route: D. L. Boger, N. Han, C. M. Tarby, C. W. Boyce, H. Cai, Q. Jin, P. A. Kitos, J. Org. Chem. 1996, 61, 4894–4912.
- [11] J. Wrobel, A. Dietrich, B. J. Gorham, K. Sestanj, J. Org. Chem. 1990, 55, 2694–2702.

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